

Lancifodilactone G: A Unique Nortriterpenoid Isolated from *Schisandra lancifolia* and Its Anti-HIV Activity

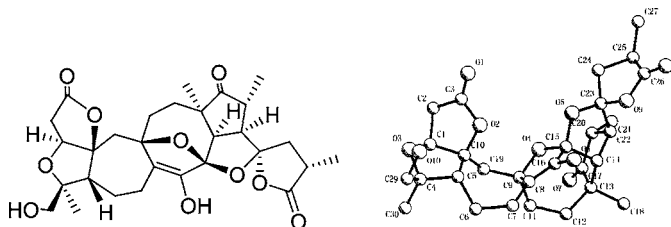
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ABSTRACT



Lancifodilactone G (1)

Lancifodilactone G (1), a novel, highly oxygenated nortriterpenoid featuring a partial enol structure and a spirocyclic moiety, was isolated from the medicinal plant *Schisandra lancifolia*. Its structure and stereochemistry were determined from extensive one- and two-dimensional NMR and mass spectral data, coupled with single-crystal X-ray analysis. Compound 1 exerted minimal cytotoxicity against C8166 cells ($CC_{50} > 200$ $\mu\text{g/mL}$) and showed anti-HIV activity with $EC_{50} = 95.47 \pm 14.19$ $\mu\text{g/mL}$ and a selectivity index in the range of 1.82–2.46.

Previous studies of *Schisandra* species have reported lignans with various beneficial pharmacological effects such as antihepatitis, antitumor, and anti-HIV activities as typical of this genus.^{1–3} Recent research showed that some triterpenoids isolated from this genus exhibited anti-HIV activities^{4,5} and inhibitory activities toward cholesterol biosynthesis.^{6–9}

Recently, we have reported the isolation and structure elucidation of several highly oxygenated nortriterpenoids with a new skeleton such as micrandilactone A,¹⁰ lancifodilactones A–F,^{11–13} and henridilactones A–D¹⁴ from the

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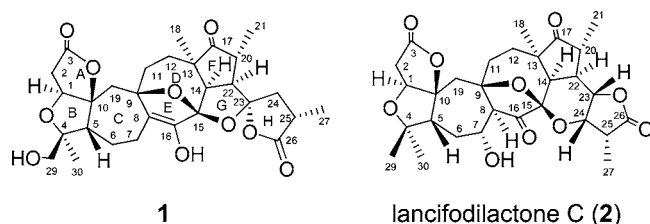
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leaves and stems of *Schisandra micrantha* A. C. Smith, *Schisandra henryi* var. *yunnanensis* A. C. Smith, and *S. lancifolia* (Rehd. et Wils) A. C. Smith, respectively. Continued phytochemical investigation of the leaves and stems of *S. lancifolia* (Rehd. et Wils) A. C. Smith led to the isolation of a novel triterpene derivative, lancifodilactone G (**1**), which possesses a highly oxidized norcycloartane skeleton similar to that of lancifodilactone C (**3**)¹² and was determined to have a partial enol structure and a spirocyclic moiety. In addition, compound **1** was tested for its cytotoxic and anti-HIV-1 activities. Described herein are the isolation, structure elucidation, and biological activity of compound **1**.



The leaves and stems of *S. lancifolia* were collected in Dali Prefecture of Yunnan Province, China, in August of 2002 and identified by Prof. Su-Gong Wu. The air-dried and powdered stems and leaves (5.7 kg) were extracted with 70% aqueous Me₂CO (4 × 15 L) at room temperature and concentrated in vacuo to give a crude extract (290 g), which was partitioned between H₂O and EtOAc. The EtOAc fraction (101 g) was subjected to column chromatography over silica gel and RP-18 repeatedly, followed by recrystallization from MeOH, yielding lancifodilactone G (**1**, 18.2 mg).

Lancifodilactone G (**1**),¹⁵ [α]_D^{25.9} +75.00 (c 0.240, pyridine), crystallized as a colorless prism and has the molecular formula C₂₉H₃₆O₁₀ as determined by analysis of ¹H, ¹³C, and DEPT NMR spectral data, which was verified by HR-ESIMS (found 567.2217, calcd 567.2206), requiring 12 degrees of unsaturation. The ¹H NMR spectrum displayed signals due to three tertiary methyls and a secondary methyl. The ¹³C NMR spectrum of **1** exhibited signals for 29 carbons, including 2 ester groups, 1 carbonyl group, 8 quaternary carbons, 6 methines (including an oxygenated one), 8 methylenes (including an oxygenated one), and 4 methyls. Comparison of ¹H and ¹³C NMR spectral data of **1** with those of lancifodilactone C (**2**)¹² showed that some characteristic signals of this kind of nortriterpenoid skeleton still existed

in compound **1** such as the proton signals at H-1 (δ 4.13, d, J = 5.3 Hz) and H-19 (δ 2.03 and 2.13, AB d, J = 15.9 Hz) and carbon signals at δ 80.8 (C-1), 97.0 (C-10), and 220.1 (C-17). All these data revealed that **1** possesses a structure partly similar to that of **2**. Considering that compound **1** and lancifodilactone C (**2**) have similar structural parts, we first tentatively established the possible structure of **1** by comparison with the NMR spectral data of **2** together with detailed analysis of two-dimensional NMR spectral data of **1**. Uncertain structure details were established by single-crystal X-ray analysis.

The differences between **1** and **2** are as follows: The methyl group at C-29 in **2** was replaced by an oxygenated methylene in **1** with a downfield shift at δ 68.2. The oxygenated methine C-7 in **2** was replaced by a methylene in **1**, which was further confirmed by HMBC correlations from H-7 at δ 2.95 (m) to C-6 and C-9. Moreover, typical signals at C-8 and C-16 in **2** were not present in the spectra of **1**. In the HMBC spectrum, correlations observed between H-7 (δ 2.95) and C-8 (δ 111.8) and C-16 (δ 148.3) and between H-6 (δ 2.12) and C-7 and C-8 suggested that there was a double bond between C-8 and C-16 in **1**. The largely downfield chemical shift of C-16 to δ 148.3 hints that there should be a hydroxyl group or an oxygen bridge connected with it. Further comparison of ¹H and ¹³C NMR data with those of **2** and analysis of two-dimensional NMR spectra of **1** (Table 1) allowed us to identify the existence of rings A–F, which led to the establishment of partial structure **1a** (Figure 1). HMBC cross-peaks (Table 1) observed from H-27 at δ 1.15 (d, J = 7.3 Hz) to C-24 and C-26 and from H-25 at δ 2.75 to C-23, C-26, and C-27, along with the proton spin system deduced from ¹H–¹H COSY correlations, H-24/H-25/H-27, established the partial structure **1b** (Figure 3). Furthermore, HMBC correlations from H-24 at δ 1.97 (d, J = 9.3 Hz, H-24 α) and 2.47 (d, J = 9.3, 13.3 Hz, H-24 β) to C-22 and from H-20 (δ 2.74) to C-23 required direct connection of C-22 with C-23 and permitted fragments **1a** and **1b** to be joined to obtain **1c** (Figure 1). Because C-15, C-16, C-23, and C-26 were quaternary carbons, it was not possible to determine the correct connections among these carbons. Luckily, a single crystal of compound **1** was achieved after repeated recrystallization, and the X-ray diffraction experiment¹⁶ was successfully performed (Figure 2), which solved the problem. X-ray diffraction indicated

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(15) Lancifodilactone G (**1**): white crystals, mp 171–172 °C; [α]_D^{25.9} +75.00 (c 0.240, MeOH); UV (MeOH) λ_{\max} (lg ϵ) 204 (3.87), 248 (2.44), 320 (2.15), 374 (1.92) nm; IR (KBr) ν_{\max} 3606, 3547, 3420, 2925, 1774, 1732, 1457, 1175, 1108, 1055, 969, 921 cm⁻¹; NMR can be found in Table 1; positive ESIMS m/z (rel intensity) 567 (100, [M + Na]⁺); HR-ESIMS found 567.2217, calcd for C₂₉H₃₆O₁₀ 567.2206.

(16) Crystallographic data for **1**: C₂₉H₃₆O₁₀, M = 544.22, monoclinic, space group $P2_1$, a = 7.348 (1) Å, b = 11.927 (1) Å, c = 15.252 (1) Å, β = 85.00 (1)°, V = 1355.4 (2) Å³, Z = 2, d = 1.379 g/cm³, crystal dimensions 0.20 × 0.20 × 0.30 mm were used for measurements on a MAC DIP-2030K diffractometer with a graphite monochromator (ω -2 θ scans, $2\theta_{\max}$ = 50.0°), Mo K α radiation. The total number of independent reflections measured was 2916, of which 2310 were observed ($|F|^2 \geq 8\sigma(F)^2$). Final indices: R_f = 0.044, R_w = 0.045 ($w = 1/\sigma(F)^2$). The crystal structure (**1**) was solved by direct methods using SHELX-86 (Sheldrick, G. M., University of Göttingen: Göttingen, Germany, 1985) and expanded using difference Fourier techniques, refined by the program and method NOMCS-DP (Lu, Y.; Wu, B. M. *Chin. Chem. Lett.* **1992**, 3, 637–640) and full-matrix least-squares calculations. Crystallographic data for the structure of **1** have been deposited in the Cambridge Crystallographic Data Centre (deposition number: CCDC 254748). Copies of these data can be obtained free of charge via the Internet at www.ccdc.cam.ac.uk/conts/retrieving.html (or from the Cambridge Crystallographic Data Centre, 12, Union Road, Cambridge CB21EZ, UK; fax: (+44) 1223-336-033; or e-mail: deposit@ccdc.cam.ac.uk).

Table 1. ^1H and ^{13}C NMR Assignments and Two-Dimensional NMR Correlations of **1**^a

position	δ_{H} (mult, J , Hz)	δ_{C} (mult)	HMBC (H–C)	NOESY (H–H)
1	4.13 (d, 5.3)	80.8 (d)	2, 3, 10, 19	2 α , 19 α , 30
2 α	2.67 (overlap)	36.0 (t)	10	1, 2 β
2 β	2.59 (d, 18.2)		1, 3, 10	2 α
3		175.1 (s)		
4		87.6 (s)		
5	2.68 (overlap)	52.7 (d)	1, 4	19 β , 29
6	2.12 (2H, m)	21.9 (t)	5, 7, 8, 16	7
7	2.95 (2H, m)	23.3 (t)	6, 8, 9, 16	6
8		111.8 (s)		
9		85.7 (s)		
10		97.0 (s)		
11 α	1.65 (m)	37.3 (t)	8, 12, 13	11 β , 12
11 β	2.33 (m)		8, 9, 12, 13, 19	11 α , 12, 19 β
12	1.80 (2H, m)	29.6 (t)	9, 11, 13, 14, 17	11 α , 11 β , 14, 18
13		50.3 (s)		
14	3.24 (d, 6.7)	57.6 (d)	13, 18, 20, 22	12, 18, 22
15		114.1 (s)		
16		148.3 (s)		
17		220.1 (s)		
18	0.99 (3H, s)	26.8 (q)	12, 13, 14, 17	12, 14, 22
19 α	2.13 (AB d, 15.9)	43.3 (t)	1, 5, 8, 9, 10	1, 19 β
19 β	2.03 (AB d, 15.9)		1, 9	5, 11 β , 19 α
20	2.74 (overlap)	43.6 (d)	17, 21, 22, 23	
21	1.27 (3H, d, 7.2)	17.8 (q)	17, 20, 22	22
22	2.68 (overlap)	53.7 (d)	14, 17, 20, 21, 23	14, 18, 21, 27
23		116.4 (s)		
24 α	1.97 (d, 9.3)	37.3 (t)	22, 23, 25, 26, 27	24 β , 27
24 β	2.47 (dd, 9.3, 13.3)		22, 23, 25, 26, 27	24 α , 25
25	2.75 (overlap)	36.0 (d)	23, 26, 27	24 β , 27
26		180.0 (s)		
27	1.15 (3H, d, 7.3)	16.4 (q)	24, 25, 26	22, 24 α , 25
29	3.63 (d, 11.6)	68.2 (t)	4, 5, 30	5, 30
30	1.15 (3H, s)	17.1 (q)	4, 5, 29	1, 29

^a Data were recorded in $\text{C}_5\text{D}_5\text{N}$ on Bruker AM-400 MHz (^1H , ^{13}C) and Bruker DRX-500 MHz spectrometers (COSY, HMBC, NOESY); chemical shifts (δ) are expressed in parts per million with reference to the most downfield signal of $\text{C}_5\text{D}_5\text{N}$ (δ 8.71 ppm) for ^1H and to the center peak of the most downfield signal of $\text{C}_5\text{D}_5\text{N}$ (δ 149.9 ppm) for ^{13}C .

that an enol structure existed between C-8 and C-16 and that a spirocyclic moiety (ring H) was located at C-23. Both are the first such examples in this series of highly oxidized novel nortriterpenoids^{10–14} and are very rare in natural products.

Regarding the stereochemistry of lancifodilactone **G** (**1**), C-29 was biogenetically β -oriented, while Me-21 and Me-

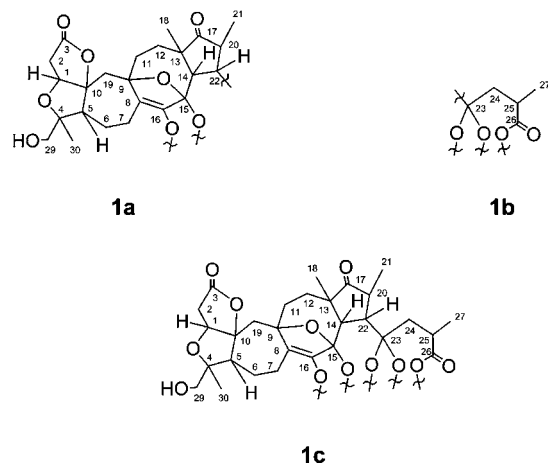


Figure 1. Structural fragments of **1**.

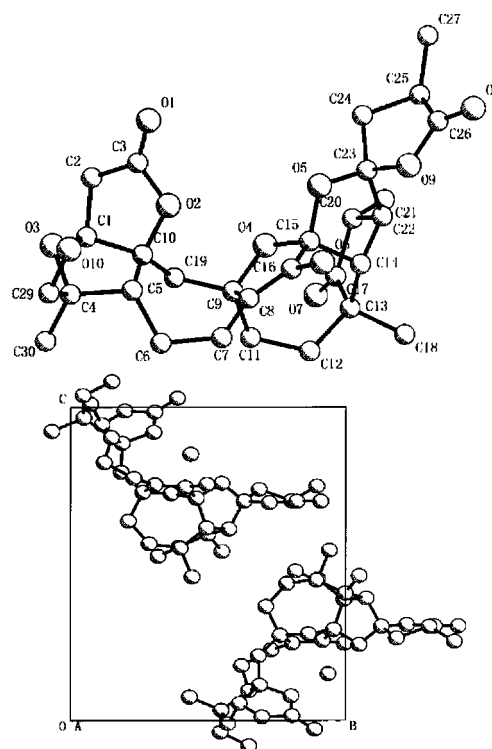


Figure 2. X-ray structure of **1** showing the relative configuration.

30 were in α -orientation. The cross-peaks observed between Me-30/H-1, H-29/H-5, H-18/H-21, H-14/H-18, and H-22/H-18 in the NOESY spectrum (Table 1) demonstrated that H-1, H-14, Me-18, and H-22 were α -oriented, while H-5 possessed β -orientation. The relative stereochemistry of the

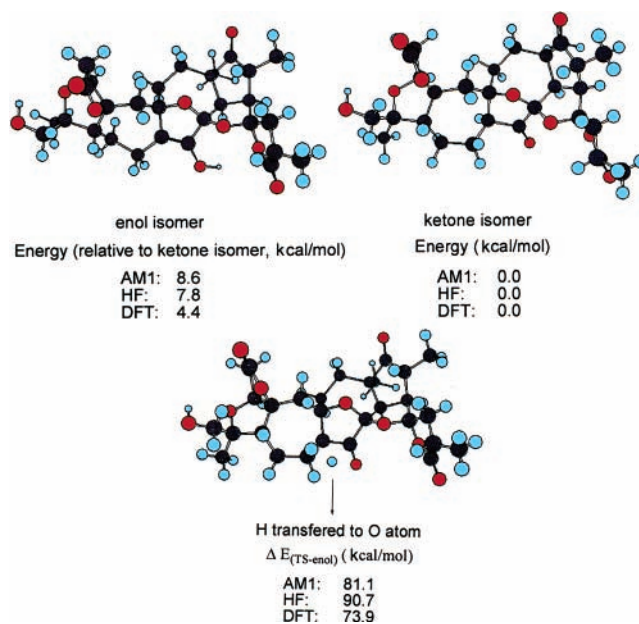
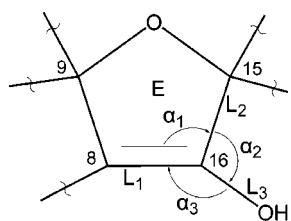


Figure 3. Relative energies of enol isomer, ketone isomer, and TS at different levels of theory.



X-ray: L₁ = 1.326, L₂ = 1.505, L₃ = 1.356
 α₁ = 110.5, α₂ = 116.3, α₃ = 133.0
 AM1: L₁ = 1.355, L₂ = 1.530, L₃ = 1.355
 α₁ = 108.6, α₂ = 123.4, α₃ = 127.9
 HF: L₁ = 1.314, L₂ = 1.509, L₃ = 1.338
 α₁ = 109.8, α₂ = 122.3, α₃ = 127.6

Figure 4. Bond lengths and bond angles in ring E obtained by computations and X-ray experiment.

four quaternary carbons C-9, C-10, C-15, and C-23 was deduced as *R*, *S*, *S*, and *S*, respectively, by an X-ray diffraction experiment.

The confirmation by X-ray experiment that lancifodilactone G exists as an enol isomer might hint that this isomer may be more stable than the ketone isomer in energy in this special molecule. However, the computed relative energies in different levels of theory showed that enol isomers are 4.4–8.6 kcal/mol (Figure 3) higher in energy than the ketone isomer,¹⁷ meaning that the enol isomer is less stable than the ketone isomer. Therefore, transition states (TSs) were investigated to explain this case. The TS computations were performed from the enol isomer to ketone at the AM1 and HF/6-31G(d,p) levels of theory; the single-point energy computations were performed using HF/6-31G(d,p)-optimized geometries at the B3LYP/6-31+G(d,p) level of theory. The calculated TS barriers are 81.1 kcal/mol using AM1, 90.7 kcal/mol using HF/6-31G(d,p), and 73.9 kcal/mol using single-point energy at the B3LYP/6-31+(d,p) (DFT method) level of theory. All barriers obtained are so high that it is almost impossible for such an isomerization to occur. The

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Table 2. Summary of Cytotoxicity and Anti-HIV-1_{IIIB} Activity of Compound **1**

compound	cytotoxicity CC ₅₀ (μg/mL) ^a	anti-HIV-1 _{IIIB}	selectivity
		activity EC ₅₀ (μg/mL)	index CC ₅₀ /EC ₅₀
lancifodilactone G (1)	>200	95.47 ± 14.19	>1.82–2.46

^a Minimal cytotoxicity against C8166 cells when CC₅₀ > 200 (μg/mL).

partial bond angles and bond lengths, which were obtained by X-ray and computations, are listed in Figure 4. These computed bond lengths and angles are close to those obtained by X-ray experiment. The above results suggest that lancifodilactone G is not an artifact. However, it remains unknown why the plant forms the enol isomer instead of the ketone structure.

Compound **1** was tested for cytotoxicity against C8166 cells (CC₅₀) using the MTT method as reported previously,¹⁸ and anti-HIV-1 activity was evaluated by the inhibition assay for the cytopathic effects of HIV-1_{IIIB} (EC₅₀)¹⁹ (Table 2). Compound **1** exerted minimal cytotoxicity against C8166 cells (CC₅₀ > 200 μg/mL) and showed anti-HIV-1_{IIIB} activity with EC₅₀ = 95.47 ± 14.19 μg mL⁻¹ and selectivity index over 1.82–2.46.

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Supporting Information Available: One- and two-dimensional NMR spectra of lancifodilactones F (**1**), the complete ref 17, and coordinates and optimized energies of computations. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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